



Reaction Assembly

| Component | Volume for 50- μ L rxn. | Final Concentration |
|-----------------------------|---------------------------------|---------------------|
| PerfeCTa qPCR SuperMix (2X) | 25 μ L | 1x |
| Forward primer | variable | 100 – 900 nM |
| Reverse primer | variable | 100 – 900 nM |
| Probe | variable | 100 – 250 nM |
| Nuclease-free water | variable | |
| Template | <u>5 – 10 μL</u> | variable |
| Final Volume (μ L) | 50 μ L | |

Note: For smaller reaction volumes (i.e. 25- μ L reactions), scale all components proportionally.

Reaction Protocol

Incubate complete reaction mix in a real-time thermal detection system as follows:

| | |
|-----------------------------|--|
| Initial denaturation: | 95°C, 2 to 3 min |
| PCR cycling (30-45 cycles:) | 95°C, 10 to 15 s |
| | 55 – 65°C, 30 to 45 s (collect and analyze data) |

Full activation of AccuStart Taq DNA polymerase occurs within 30 seconds at 95°C. Initial denaturation times greater than 3 minutes are usually not required. However, amplification of genomic DNA or supercoiled plasmid DNA targets may benefit from a prolonged initial denaturation step (5-10 min) to fully denature and fragment the template. This minimizes the potential for renaturation of long fragments and/or repetitive sequence regions that can impair replication of the target sequence by the PCR process.

Some primer sets may require a 3-step cycling protocol for optimal performance. Optimal annealing temperature and time may need to be empirically determined for any given primer set. A 68°C extension step of 30 seconds is suitable for most applications. Amplicons greater than 200 bp may require longer extension times.

Quality Control

Kit components are free of contaminating DNase and RNase. PerfeCTa qPCR SuperMix is functionally tested in qPCR. Kinetic analysis must demonstrate linear resolution over six orders of dynamic range ($r^2 > 0.995$) and a PCR efficiency $> 90\%$.

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